Structural basis for disassembly of katanin heterododecamers

Stanley Nithianantham, Francis J. McNally, and Jawdat Al-Bassam

From the Department of Molecular Cellular Biology University of California, Davis, California 95616

Edited by Vela M. Fowler

The reorganization of microtubules in mitosis, meiosis, and development requires the microtubule-severing activity of katanin. Katanin is a heterodimer composed of an ATPase associated with diverse cellular activities (AAA) subunit and a regulatory subunit. Microtubule severing requires ATP hydrolysis by katanin’s conserved AAA ATPase domains. Whereas other AAA ATPases form stable hexamers, we show that katanin forms only a monomer or dimers of heterodimers in solution. Katanin oligomers consistent with hexamers of heterodimers or heterododecamers were only observed for an ATP hydrolysis-deficient mutant in the presence of ATP. X-ray structures of katanin’s AAA ATPase in monomeric nucleotide-free and pseudo-oligomeric ADP-bound states revealed conformational changes in the AAA subdomains that explained the structural basis for the instability of the katanin heterododecamer. We propose that the rapid dissociation of katanin AAA oligomers may lead to an autoinhibited state that prevents inappropriate microtubule severing or that cyclical disassembly into heterodimers may critically contribute to the microtubule-severing mechanism.

This work was supported by National Institutes of Health Grants GM110283 (to J. A.-B.) and GM079421 (to F. J. M.). The authors declare that they have no conflicts of interest with the contents of this article. The content is solely the responsibility of the authors and does not necessarily represent the official views of the National Institutes of Health.

This article contains Figs. 1–5

The atomic coordinates and structure factors (codes 6B5C and 6B5D) have been deposited in the Protein Data Bank (http://www.pdb.org/).

1 To whom correspondence should be addressed. E-mail: jawdat@ucdavis.edu.

2 The abbreviations used are: MT, microtubule; MIT, microtubule-interacting and trafficking; VPS4, vacuolar protein sorting-associated protein 4; con80, conserved p80; NSF, N-ethylmaleimide–sensitive fusion; SEC, size-exclusion chromatography; SEC-MALS, SEC-coupled multivariate light scattering; AMPNP, S-adenosyl-L-β−γ-imidodiphosphate; SAD, single-wavelength anomalous dispersion; NBD, nucleotide-binding domain; HBD, 4-helix bundle domain; N-Hlx, N-terminal helix; C-Hlx, C-terminal helix; RMSD, root-mean-square deviation; TEV, tobacco etch virus; GST, glutathione S-transferase; PDB, Protein Data Bank.

Microtubules (MTs)2 are dynamic cytoskeleton polymers that are essential force generators that organize the cytoplasm during cell division, development, and morphogenesis. The stability of the MT polymer is mediated by longitudinal and lateral interfaces between αβ-tubulins polymerized in the MT lattice. Diverse classes of MT regulators promote the polymerization and depolymerization of dynamic MTs by binding αβ-tubulins at their ends. However, in contrast to these regulators, MT-severing enzymes destabilize MTs by binding along MT lattice sites to generate several new MTs. The MT-severing proteins include the closely related AAA ATPases katanin, spastin, and fidgetin, which are conserved across protzoa, plants, and metazoaos (1). MT-severing enzymes carry out essential MT regulatory functions in many cellular settings in which MT are involved. During mitosis and meiosis, they activate MT disassembly (2, 3). During neuronal development, they are essential to the release of new MTs after nucleation (4), and during cell motility they regulate MT formation in cilia or flagella (5). Defects in human katanin and spastin lead to neurological disorders such as microlissencephaly (6, 7) and hereditary spastic paraplegia (8), respectively.

Katanin, first purified from sea urchin eggs, is composed of a catalytic subunit, termed p60, and a regulatory subunit, termed p80 (9). p60 katanin is composed of an N-terminal microtubule-interacting and trafficking (MIT) domain (10) followed by a 50–70-residue linker and a highly conserved C-terminal AAA ATPase domain. p80 katanin exists in multiple forms, which either include or exclude a large N-terminal β-propeller or WD-40 domain followed by a core conserved 280-residue helical bundle region termed conserved p80 (con80) (2) (Fig. 1A). Both the p60 (MIT and AAA domains) and p80 (con80 with or without WD-40) domains are essential for the MT-severing functions. p60 and p80 katanin form a complex through the con80-MIT domains, the structure of which reveals a helical assembly that is responsible for binding MTs and recruiting MT regulatory factors to sites of MT lattice deformation (2). p60 and p80 katanin co-purify from many cell types (9, 11), and p60 and p80 mutants have identical phenotypes in many organisms (5, 12), indicating that a complex of p60 and p80 is the relevant physiological complex.

Most AAA ATPases are thought to act as hexameric ring complexes. For example, N-ethylmaleimide–sensitive fusion (NSF) is greater than 90% hexameric in solution throughout a concentration range of 0.2 to 10 μM in ATP (13). X-ray or cryo-EM structures of several other WT AAA ATPases have revealed open or closed hexameric rings (14–16). In contrast, members of one subfamily of AAA ATPases are reported to be predominantly monomeric or dimeric in solution. This subfamily includes the MT-severing proteins katanin, spastin, and fidgetin, as well as Vps4 and MSP1, which disassemble nontubulin substrates (17–21). The WT form of each of these ATPases has most often been shown to be monomeric in solution at low concentrations. In contrast, each of these proteins has been reported to form a stable hexamer at low concentration when ATP hydrolysis is blocked by the mutation of a con-
erved Glu (E) in the Walker B motif to Gln (Q) (17–21). Structural studies have revealed that the ATP hydrolysis-deficient Walker B mutant of p60 katanin can form a right-handed hexameric spiral with all six protomers bound by ATP or a closed hexameric ring with five ATP-bound protomers and one nucleotide-free protomer (22). However, earlier fluorescence resonance energy transfer (FRET) experiments indicate that oligomers of WT p60 katanin could be detected only in the presence of both a microtubule substrate and a nonhydrolyzable ATP analog (19). Thus, the structural basis for katanin’s disinclination to form hexamers in the absence of a microtubule has remained unclear.

Here, we reconstituted katanin complexes of full-length p60 with core con80 domains using the Caenorhabditis elegans MEI-1/MEI-2 and human KATNAL1/KATNB1 forms. We show that p60/p80 is a heterodimer or heterotetramer with a poor capacity for oligomerization into oligomers, most consistent with heterododecamers. The crystal structures for katanin’s AAA ATPase in the ADP-bound and nucleotide-free states reveal a substantial conformational transition that could explain katanin’s poor propensity for oligomerization.

Results

Catalytically active p60/p80 katanin is composed predominantly of heterodimers or heterotetramers in solution

To examine the oligomerization properties of p60/p80 katanin, we first purified complexes of full-length C. elegans MEI-1 with full-length MEI-2 (termed MEI-1/MEI-2), and full-length human KATNAL1 with the con80 domain (residues 411–655) of human KATNB1 (termed KATNAL1/B1-con80) (Figs. 1A and S1, A and B). MEI-1 and KATNAL1 are 51.7- and 55.4-kDa proteins, respectively, whereas MEI-2 and KATNB1-con80 are 31.5- and 26.7-kDa proteins, respectively. Thus, isolated heterodimers of MEI-1/MEI-2 or KATNAL1/B1-con80 are roughly 83 kDa in mass. KATNAL1/B1-con80 complexes were studied using a Superose 6 size-exclusion chromatography (SEC) column at katanin concentrations ranging from 5 to 50 μM in the presence of 1 mM ATP (Figs. 1, B and D, and S2A). Two overlapping peaks eluted earlier at higher concentrations, indicating a concentration-dependent increase in Stokes radius as reported previously for MEI-1/MEI-2 complexes (23). SDS-PAGE of the peak fractions indicated that the subunit masses and the stoichiometry of KATNAL1 to B1-con80 were the same in all the peak fractions at all concentrations (Fig. 1, D and E, and S2A). To test whether this increase in Stokes radius was because of an increase in mass due to oligomerization of heterodimers, 10–50 μM KATNAL1/B1-con80 complexes were analyzed by SEC-coupled multiangle light scattering (SEC-MALS) in the presence of 1 mM ATP using a Superdex 200 column (Fig. 1C and Table 1). The two peaks of katanin complexes were more clearly resolved using a Superdex 200 SEC column. SEC-MALS measurements indicated a mass for the faster eluting complex that was most consistent with a dimer of heterodimers (heterotetramer) and a mass for the slower eluting complex that was most consistent with a heterodimer (Fig. 1C). Strikingly, the SEC-MALS–measured masses did not change in the range of 10 to 50 μM katanin (Table 1) even though the Stokes radii increased in this concentration range (Fig. 1, B and C). C. elegans MEI-1/MEI-2 eluted as a single peak in the absence of ATP (Fig. S1A). SDS-PAGE and quantitation of MEI-1 to MEI-2 molar ratios indicated a 1:1 molar ratio similar to human KATNAL1/B1-con80 complexes (Fig. S1D). MALS measurements of MEI-1/MEI-2 indicated a mass intermediate between that of a dimer of heterodimers and that of a trimer of heterodimers (MEI-1/MEI-2 heterodimer, 83 kDa; and SEC-MALS–measured mass, 216 kDa) (Fig. S1, C–E and Table 1), suggesting a possible equilibrium but no assembly into a hexamer of heterodimers. Our SEC-MALS results indicate that WT KATNAL1/B1-con80 does not assemble beyond dimers of heterodimers at concentrations up to 50 μM. For WT KATNAL1/B1-con80, masses were identical in the absence (dimers of heterodimers measured mass, 164–169 kDa; predicted mass, 164.2 kDa) (Figs. 1F and S2B) or in the presence (Figs. 1G and S2B and Table 1) of ATP. Previous studies of katanin (19, 22) and spastin (18) indicate that a Glu → Gln mutation in the Walker B motif stabilized a hexameric assembly only in the presence of ATP. Indeed, an E308Q variant of KATNAL1/B1-con80 had a mass most consistent with a heterododecamer or hexamer of heterodimers (measured mass, 505 kDa; and predicted mass, 492 kDa) only in the presence of ATP (Fig. 1G and Table 1). In the absence of ATP, KATNAL1 (E308Q)/B1-con80 eluted as two peaks with masses consistent with heterodimer and dimer of heterodimers (heterotetramer), similar to WT KATNAL1/B1-con80 with or without ATP. Strikingly, the Stokes radius of a KATNAL1 (E308Q)/B1-con80 hexamer of heterodimers (heterododecamer) was nearly identical to that of a WT dimer of heterodimers (heterotetramer) at the same concentration, suggesting that the dimer of heterodimers has an elongated conformation (Fig. 1G). SDS-PAGE and quantitation of molar ratios for KATNAL1 WT or E308Q compared with KATNB1-con80 show that they are present at 1:1 molar stoichiometry and that there are no differences in subunit compositions in the different states (Fig. 1E and Fig. S2). Previous work estimates the in vivo concentration of katanin at 20–50 nm (11). Our results thus indicate that WT katanin does not form hexamers of heterodimers (heterododecamers) at concentrations 1000 times higher than physiological concentrations and suggest that, in solution, WT katanin is in an autoinhibited state in the presence or absence of nucleotide. Furthermore, ATP is required for the oligomerization of KATNAL1 (E308Q)/B1-con80, suggesting that dissociation of the nucleotide returns katanin to its autoinhibited state (Fig. 1, G and H).

Crystal structure of MEI-1’s AAA ATPase in the ADP-bound state reveals a pseudo-hexameric left-handed spiral assembly

To understand the structural basis for nucleotide-driven katanin oligomerization, we aimed to determine the crystal structure of katanin with the nonhydrolyzable ATP analog AMPPNP. Although we crystallized the full-length C. elegans p60 katanin in complex with the con80 domain of p80, MEI-1/MEI-2 crystals contained only the AAA ATPase domains and bound ADP molecules. The MIT/Con80 domains degraded during crystallization, likely because of the prolonged incubation (14 days) required for crystallization (Fig. S2F). The AMP-PNP presumably either hydrolyzed during crystallization or...
Katanin nucleotide-free state reveals disassembly mechanism

(A) Katanin nucleotide-free state reveals disassembly mechanism

(B) Elongated conformations

(C) Equilibrium between heterodimers and heterotetramers

(D) SDS-PAGE of the peak fractions at concentrations ranging from 5 to 50 μM

(E) KATNAL1/B1-con80 (molar ratio)

(F) No nucleotide Wt complex E-Q complex

(G) 1 mM ATP Wt complex E-Q complex

(H) ATP + MT → Heterotetramers → Heterododecamer

KATNAL1/B1 KATNAL1/B1-con80 MEI-1/MEI-2

KATNAL1 KATNB1

5 μM 15 μM 25 μM 50 μM

5 kDa 15 kDa 25 kDa 50 kDa

Retention Volume (mL)

Normalized dfN

Normalized dfN

Retention Volume (mL)

Molar Mass (g/mol)

Retention Volume (mL)

Molar Mass (g/mol)

Retention Volume (mL)

Molar Mass (g/mol)
never exchanged with nucleotide bound during bacterial expression. Crystals of C. elegans MEI-1/MEI-2 formed in the presence of ADP in the space group P6_3, and diffracted to 3.1 Å resolution. Phase information was determined by single-wavelength anomalous dispersion (SAD) using selenomethionine-substituted protein (see “Experimental procedures”). The refined 3.1Å structure includes only part of the AAA-MIT linker region (residues 164–171) and the MEI-1 AAA domain starting at residue 172 and extending to residue 468 (Fig. 2, A and B, and Table 2) in which three flexible loops are disordered (see “Experimental procedures”). The structure reveals an AAA ATPase fold with ADP (Fig. 2B) that is similar to the recently reported X-ray structure of the E to Q ATP hydrolysis deficient mutant of sulfate-bound MEI-1 AAA (22). Briefly, as with all AAA structures, MEI-1 AAA consists of a larger nucleotide-binding domain (NBD) and a smaller 4-helix bundle domain (HBD). The NBD is in an α/β Rossman fold consisting of a 5-stranded parallel β-sheet (β1–β5) sandwiched between nine α-helices (α2–α10). The NBD cradles an ADP molecule via Walker A and Walker B motifs (Fig. 2, E and F). The HBD is composed of a central α-helix bound orthogonally by four antiparallel helices (α11–α13 and α16) with its sensor II motif. Two additional short helices (α14–α15) were inserted between α13 and α16. This insertion is replaced by the β-domain in the Vps4 AAA ATPase (21). The MEI-1 AAA structure reveals two highly conserved katanin expansion AAA ATPase segments as seen previously (22), which we term the N-terminal (α1, N-Helix) and C-terminal (α17, C-Helix) helices, respectively (Figs. 2, A and B, and S3). The N-Helix consists of a single-turn helix bound along one side of the NBD followed by a linker and a three-turn helix bound along the opposing face of the NBD (Fig. 2B). The C-Helix stabilizes the NBD and HBD junction, connected by a short loop that we term the hinge. Like the Glu → Gln mutant structure (22), our WT MEI-1 AAA structure reveals the conformation of the AAA subdomains in a pseudo-oligomeric assembly state. In this structure, the protomers are arranged along a pseudo-hexameric left-handed spur assembly with one subunit in the asymmetric unit (Fig. 2C). The projection of this structure parallel to this crystallographic screw axis reveals a pseudo-hexameric left-handed spiral assembly with a 14-Å translation between two adjacent subunits (Fig. S4, A and B). In this conformation, the NBD and ADP of one AAA subunit is bound by an HBD via the sensor II motif with a 1112-Å buried surface/monomer that might reflect the functional contacts of a physiologically relevant oligomer (Fig. 2, C, E, and F). The expansion N-Helix and the C-Helix line opposite sides of this left-handed katanin spiral. ADP was clearly defined in our MEI-1 structure, suggesting a possible relationship between ADP binding and the left-handed spiral assembly. Alternatively, this ADP-bound structure could represent a monomer that is competent to assemble into a right-handed hexameric spiral or closed ring (22). Clearly, neither a continuous spiral nor a hexameric ring formed in our SEC-MALS experiments on WT KATNAL1/B1-con80, and therefore we sought a possible mechanism to prevent assembly in solution.

**Crystal structure of KATNAL1’s AAA ATPase in the nucleotide-free state reveals a monomer**

We also attempted to determine the structure of human KATNAL1 (E308Q)/B1-con80, and crystals grew in the space group P2_1_2_1. We observed the degradation of the MIT/con80 domains during crystallization similar to MEI-1/MEI-2, likely because of the prolonged incubation time required for crystals to form. We determined the X-ray structure of the KATNAL1

### Table 1

<table>
<thead>
<tr>
<th>Protein</th>
<th>Conc.</th>
<th>Nucleotide</th>
<th>Hetero-dimer^a,b</th>
<th>Heterotetramer^c,d</th>
<th>Heterododecamer^e</th>
<th>Subunit molar ratio (SDS-PAGE)</th>
</tr>
</thead>
<tbody>
<tr>
<td>WT KATNAL1/B1-con80</td>
<td>10</td>
<td>ATP</td>
<td>93.81 ± 5.52</td>
<td>166.51 ± 6.00</td>
<td>505.40 ± 4.02</td>
<td>0.903 ± 0.067</td>
</tr>
<tr>
<td>WT KATNAL1/B1-con80</td>
<td>25</td>
<td>ATP</td>
<td>98.77 ± 4.82</td>
<td>164.60 ± 5.40</td>
<td>505.40 ± 4.02</td>
<td>0.910 ± 0.047</td>
</tr>
<tr>
<td>WT KATNAL1/B1-con80</td>
<td>50</td>
<td>ATP</td>
<td>87.88 ± 3.30</td>
<td>169.01 ± 2.20</td>
<td>505.40 ± 4.02</td>
<td>0.879 ± 0.056</td>
</tr>
<tr>
<td>WT KATNAL1/B1-con80</td>
<td>20</td>
<td>ATP</td>
<td>84.23 ± 6.70</td>
<td>160.80 ± 4.40</td>
<td>505.40 ± 4.02</td>
<td>0.952 ± 0.085</td>
</tr>
<tr>
<td>WT KATNAL1/B1-con80</td>
<td>20</td>
<td>ATP</td>
<td>85.98 ± 7.80</td>
<td>166.60 ± 3.60</td>
<td>505.40 ± 4.02</td>
<td>0.925 ± 0.043</td>
</tr>
<tr>
<td>KATNAL1 (E308Q)/B1-con80</td>
<td>25</td>
<td>ATP</td>
<td>90.30 ± 1.21</td>
<td>171.60 ± 4.12</td>
<td>216.50 ± 7.10c</td>
<td>0.916 ± 0.058</td>
</tr>
<tr>
<td>KATNAL1 (E308Q)/B1-con80</td>
<td>50</td>
<td>ATP</td>
<td>90.30 ± 1.21</td>
<td>171.60 ± 4.12</td>
<td>216.50 ± 7.10c</td>
<td>0.923 ± 0.091</td>
</tr>
</tbody>
</table>

^a Calculated mass for KATNAL1/B1-con80 heterodimer: 82.1 kDa (55.4 ± 26.7 = 82.1 kDa).
^b Calculated mass for a MEI-1/MEI-2 heterodimer: 83.2 kDa (51.7 ± 31.5 = 83.2 kDa).
^c Calculated mass for a heterotetramer (dimer of KATNAL1/B1-con80 heterodimers): 164.2 kDa.
^d Calculated mass for a heterotetramer (dimer of MEI-1/MEI-2 heterodimers): 166.4 kDa.
^e Calculated mass for a heterododecamer (hexamer of KATNAL1/B1-con80 heterodimers): 492.6 kDa.
^f Calculated mass for a heterohexamer (trimer of MEI-1/MEI-2 heterodimers): 249.6 kDa.

Figure 1. Domain structures and biochemical characterization of katanin. A, schematic representation of Homo sapiens katanin, a heterodimer of p60 and p80 proteins (KATNAL1/KATNB1) consisting of MIT (blue), AAA ATPase (red, N-Helix; light green, NBD; cyan, 4-helix bundle domain or HBD; orange, C-Helix), β-propeller or WD-40 (green), and conserved p80 or con-80 (light purple) domains, respectively. The KATNAL1/B1-con80 and C. elegans MEI-1/MEI-2 structures are shown below. B, SEC-based analysis of WT KATNAL1/B1-con80 at concentrations ranging from 5 to 50 μM in the presence of 1 mM ATP using size-exclusion chromatography (Superose 6 column). Note that the complex migrates more slowly at lower concentrations, suggesting that KATNAL1/B1-con80 is in equilibrium. C, SEC-MALS of WT KATNAL1/B1-con80 at the same concentrations in the presence of 1 mM ATP using a Superdex 200 column shows monomer to dimers of heterodimers (heterodimers and heterotetramers), revealing that the complex is in equilibrium. D, SDS-PAGE of the peak fractions (red and black marks) of WT KATNAL1/B1-con80 complexes as shown in Figs. 2A, A and B, F, SEC-MALS of WT (black) and E308Q mutant (red) of KATNAL1/B1-con80 reveals that the complexes are in a heterodimer–heterotetramer equilibrium in the absence of 1 mM ATP. G, SEC-MALS curves reveal that KATNAL1 (E308Q)/B1-con80 (red) forms a heterododecameric assembly in the presence of 1 mM ATP but only a heterotetrameric assembly with WT KATNAL1/B1-con80 (black). H, schematic reaction scheme of katanin p60/p80 oligomerization and the role of ATP binding and hydrolysis in this process.
Katanin nucleotide-free state reveals disassembly mechanism

A

B

C

D

E

F
The last three or four conserved residues of C-terminal structure, this structure is not in a spiral-like conformation (Fig. 2). Therefore, we analyzed the density content in the nucleotide-binding pocket including the Walker A and B motifs in a conformation similar to those in the ADP-bound intermediate and our monomeric structure represents a transient nucleotide-free state might appear to be inconsistent with the earlier X-ray structures of MEI-1 AAA (22) and C. elegans MEI-1 (47% identity and 85% similarity) or might reflect differences between the ADP-bound and nucleotide-free states during an ATP hydrolysis cycle. The suggestion that the left-handed spiral assembly of katanin reflects a transient ADP-bound intermediate and our monomeric structure represents a transient nucleotide-free state might appear to be inconsistent with the earlier X-ray structures of MEI-1 AAA (22) and spastin (18, 24). Katanin and spastin AAA domains crystallized in a left-handed spiral arrangement with no nucleotide bound. Therefore, we analyzed the density content in the nucleotide pockets from the deposited maps for two spastin AAA and MEI-1 (Glu → Gln) structures to our MEI-1 AAA-ADP structure (18, 22, 24) (Fig. S6). Density maps of the ATP nucleotide-binding pockets revealed the presence of chloride ion or sulfate molecules in the published katanin and spastin structures. The absence of nucleotide or any potential mimic molecule (sulfate or chloride ion) in the crystals reveals that KATNAL1 (E308Q) AAA is a monomer (Fig. 1D and Table 1). Thus, this structure likely represents the autoinhibited state of the katanin AAA domain.

The differences between our ADP-bound MEI-1 structure and our nucleotide-free KATNAL1 structure might be because of sequence divergence between human KATNAL1 and C. elegans MEI-1 (47% identity and 85% similarity) or might reflect differences between the ADP-bound and nucleotide-free states during an ATP hydrolysis cycle. The suggestion that the left-handed spiral assembly of katanin reflects a transient ADP-bound intermediate and our monomeric structure represents a transient nucleotide-free state might appear to be inconsistent with the earlier X-ray structures of MEI-1 AAA (22) and spastin (18, 24). Katanin and spastin AAA domains crystallized in a left-handed spiral arrangement with no nucleotide bound. Therefore, we analyzed the density content in the nucleotide pockets from the deposited maps for two spastin AAA and MEI-1 (Glu → Gln) structures to our MEI-1 AAA-ADP structure (18, 22, 24) (Fig. S6). Density maps of the ATP nucleotide-binding pockets revealed the presence of chloride ion or sulfate molecules in the published katanin and spastin structures. The

Table 2
Crystallographic statistics

<table>
<thead>
<tr>
<th>Statistics</th>
<th>KATNAL1 (E308Q) AAA</th>
<th>KATNAL1 (E308Q) AAA gold derivative (Peak)</th>
<th>MEI-1 AAA (SeMet peak)</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Data collection</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Resolution range (Å)</td>
<td>61.19–2.40 (2.53–2.40)</td>
<td>60.88–3.90 (4.11–3.90)</td>
<td>85.59–3.10 (3.27–3.10)</td>
</tr>
<tr>
<td>Space group</td>
<td>P2₁ 2₁ 2₁</td>
<td>P2₁ 2₁ 2₁</td>
<td>P₆₆</td>
</tr>
<tr>
<td>Wavelength (Å)</td>
<td>0.9792</td>
<td>1.0388</td>
<td>0.9791</td>
</tr>
<tr>
<td>Unit cell (Å): a, b, c</td>
<td>40.10, 61.19, 117.62</td>
<td>40.30, 60.88, 118.70</td>
<td>98.83, 98.83, 75.08</td>
</tr>
<tr>
<td>Total number of observed reflections</td>
<td>7,359/1737</td>
<td>1,1699 (1763)</td>
<td>3,757/1 (5592)</td>
</tr>
<tr>
<td>Unique reflections</td>
<td>1,1932 (1713)</td>
<td>2897 (414)</td>
<td>7653 (1118)</td>
</tr>
<tr>
<td>Average mosaicty</td>
<td>0.71</td>
<td>0.45</td>
<td>0.85</td>
</tr>
<tr>
<td>Anomalous Multiplicity</td>
<td>2.3 (2.3)</td>
<td>2.4 (2.4)</td>
<td>4.9 (5.0)</td>
</tr>
<tr>
<td>Anomalous completeness (%)</td>
<td>6.2 (4.3)</td>
<td>92.9 (95.6)</td>
<td>94.7 (94.6)</td>
</tr>
<tr>
<td>Completeness (%)</td>
<td>99.9 (99.4)</td>
<td>98.8 (99.2)</td>
<td>99.6 (100.0)</td>
</tr>
<tr>
<td>Rmerge (%)</td>
<td>0.094 (0.569)</td>
<td>0.18 (0.42)</td>
<td>0.076 (0.624)</td>
</tr>
<tr>
<td><strong>Structure refinement</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Rwork (%)</td>
<td>0.22 (0.28)</td>
<td>0.21 (0.24)</td>
<td>0.26 (0.35)</td>
</tr>
<tr>
<td>Rfree (%)</td>
<td>0.25 (0.32)</td>
<td>0.03 (0.03)</td>
<td>0.70</td>
</tr>
<tr>
<td>Molecules/asymmetric unit</td>
<td>1</td>
<td>1</td>
<td>1</td>
</tr>
<tr>
<td>Number of atoms</td>
<td>2385</td>
<td>2229</td>
<td>2202</td>
</tr>
<tr>
<td>Protein atoms</td>
<td>2319</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Ligand atoms</td>
<td>13</td>
<td>27</td>
<td></td>
</tr>
<tr>
<td>RMS bond lengths (Å)</td>
<td>0.002</td>
<td>0.003</td>
<td>1.2</td>
</tr>
<tr>
<td>RMS bond angles (°)</td>
<td>0.51</td>
<td>0.70</td>
<td>0.4</td>
</tr>
<tr>
<td>Ramachandran favored (%)</td>
<td>97.5</td>
<td>94.2</td>
<td>5.1</td>
</tr>
<tr>
<td>Ramachandran allowed (%)</td>
<td>2.1</td>
<td>5.1</td>
<td>0.7</td>
</tr>
<tr>
<td>Ramachandran outliers (%)</td>
<td>0.4</td>
<td>0.7</td>
<td></td>
</tr>
<tr>
<td>Clash score</td>
<td>5.0</td>
<td>8.0</td>
<td></td>
</tr>
<tr>
<td>Mean B-values (Å²)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Overall</td>
<td>54.33</td>
<td></td>
<td>99.20</td>
</tr>
<tr>
<td>Macromolecules</td>
<td>54.27</td>
<td></td>
<td>99.36</td>
</tr>
<tr>
<td>Ligands</td>
<td>78.13</td>
<td></td>
<td>86.36</td>
</tr>
<tr>
<td>Water</td>
<td>51.17</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

* Rmerge = Σi=1NΣn=1N(Ii(hkl)−<I(hkl)>)/Σi=1NΣn=1N(I(hkl)).
β-phosphate of ADP in our MEI-1 structure resides in a position similar to the anion in the other structures (Fig. S6, A–D). Thus, we suggest that these molecules may mimic ADP nucleotide-bound states in the previously published katanin and spastin structures, which all form spiral assemblies. We further suggest that only our KATNAL1 (E308Q) AAA structure is in a true monomeric, nucleotide-free conformation. The human KATNAL1 autoinhibited state might be more stable compared with that of MEI-1 or spastin.

Structural comparisons of katanin’s AAA ATPase in the ATP, ADP, and nucleotide-free states reveal a conformational transition that inhibits oligomerization

We compared the MEI-1 AAA ATPase structure in its pseudo-hexameric ADP state to the KATNAL1 (E308Q) AAA structure in its monomeric nucleotide-free state by superimposing the AAA domains in the two structures. The AAA ATPase structures show a root-mean-square deviation (RMSD) of 1.50 Å (Cα positions) (Fig. 3, A and B). The comparison reveals that the NBD fold becomes decompressed because of conformational changes in the N-Hlx and C-Hlx and a refolding of the HBD domain. The human KATNAL1 autoinhibited state might be more stable compared with that of MEI-1 or spastin.
Katanin nucleotide-free state reveals disassembly mechanism

Nearly orthogonal orientation to α13. The HBD (α11–α13 and α16) rotates 9° closer to the NBD. In this conformation, the HBD fold reorganizes and changes orientation, leading to clockwise subdomain rotation (α15 and α14) (Fig. 3, A and B). The NBD in the monomeric structure shows decompression-like transitions compared with its fold in the pseudo-oligomeric structure, including a 12° rotation of the α2-α3 helix-loop-helix elements, and a 10° α6 rotation. These NBD transitions involve elements that mediate NBD–NBD interfaces in the pseudo-oligomeric state.

To find out the difference between the ATP and nucleotide-free states, we built a homology model of a KATNAL1 AAA-ATP state structure based on the previously published cryo-EM structure of the MEI-1 Glu → Gln mutant in an ATP-bound hexameric state (22). We calculated those helical movements mentioned previously by superimposing the two structures (RMSD 2.43 Å; Ca positions), revealing that the rotational motions of these helices in the ATP state are amplified into slightly larger motions. The rotation angles are indicated in Fig. 3C. These findings suggest that the conformation of katanin is strictly dependent on ATP or ADP binding. The overlay of the KATNAL1 (E308Q) AAA monomeric X-ray structure onto the MEI-1 AAA spiral assembly reveals that the HBD in the KATNAL1 (E308Q) AAA structure is not compatible with its docked state onto the NBD in the MEI-1 AAA structure. The conformation of the HBD in the monomeric state likely inhibits AAA pseudo-oligomeric assembly by interfering with its NBD interface (Figs. 3D and 4). The monomeric nucleotide-free conformation is also incompatible with the hexameric right-handed spiral assembly of MEI-1 EQ AAA, the hexameric closed-ring assembly of MEI-1 EQ AAA, and the hexameric ring assembly of Vps4 in the substrate-bound state (22, 25) (Fig. 5). The RMSD values between KATNAL1 (E308Q) AAA and various katanin AAA ATPase ortholog structures are given in Table 3, revealing that ADP-state MEI-1 AAA and nucleotide-free–state KATNAL1 (E308Q) AAA structures are in different conformations (Fig. 6). The nucleotide-free structure of KATNAL1 thus might reveal the structural basis for autoinhibition.

Mapping mei-1 mutations onto the full p60 katanin structural model

Using the sequenced mei-1 loci (26), we mapped the locations of these point mutations onto the full-length p60 structure, including both AAA ATPase and MIT domains. The MIT domain of MEI-1 was modeled based on the recent crystal structure of Mus musculus p60N/p80C katanin complex (2). The map of the mei-1 mutations is shown in Fig. 7, suggesting that the majority of the residues fall in the NBD. These mutations likely lead to its misfolding or interfere in its ability to bind ATP. Three of these mei-1 mutations (sb3, sb23, ct103) lie at the junction of the NBD and HBD subdomains. One mei-1 mutation (ct89) lies in the HBD tip region, which we hypothesized is important for self-regulated monomer-to-oligomer conformational change. A single mei-1 mutation (ct99) lies in the MIT domain and likely destabilizes the MIT structure and interferes with MT binding. Only a single mei-1 mutation (ct84) is exposed on the surface, suggesting that it may interfere with NBD–NBD assembly as formed in the katanin oligomers.

Discussion

Here, we studied the biochemical and structural mechanisms for assembly and disassembly of katanin oligomers and revealed a role for ATP hydrolysis in this process. Our SEC-MALS analysis clearly demonstrated that human KATNAL1/B1-con80 does not assemble beyond a dimer of heterodimers (heterotramer), even at concentrations up to 50 μM. This result is consistent with calculations of native molecular weight for sea urchin p60/p80 katanin (9, 27) and is supported by SEC-MALS and SEC results suggesting that C. elegans MEI-1/MEI-2, which we show, is an equilibrium between a dimer and a trimer of heterodimers. Our results are in contrast with analytical ultracentrifugation results indicating that C. elegans MEI-1 alone hexamerizes at high concentration (22) and SEC results indicating MEI-1/MEI-2 can form hexamers of heterodimers (22, 23). The in vivo concentration of katanin p60/p80 in HeLa cells was estimated to be 20–50 nm using an antibody that cross-reacts with both KATNA1 and KATNAL1 (11). Thus, human katanin’s poor propensity for oligomerization may have an important regulatory role in restricting its activities to cellular locations like spindle poles, where it is concentrated and co-localized with positive regulators like ASPM1 (2). Alternatively, the dissociation of katanin heterodimers upon the release of ADP from heterododecamers might be an essential step in the microtubule-severing cycle. A recent model for ESCRT-III disassembly by VPS4 suggests a role for a treadmill hexameric spiral in which a new ATP-bound VPS4 is added at one end and ATP hydrolysis results in dissociation of a VPS4 at the other end (28). If katanin functioned in a similar way, both the right-handed spiral and closed-ring structures determined by Zehr et al. (22) would require complete dissociation of the “oldest” protomer before a new ATP-bound protomer could be added. The nucleotide-free state KATNAL1 AAA structure presented here is incompatible with all the possible oligomeric assemblies such as right-handed hexameric spiral, hexameric ring, and left-handed pseudo-hexameric spiral.

The left-handed ADP-MEI-1 spiral X-ray structure has been discounted previously as a crystal-packing artifact, as it could form endless oligomers, which have not been detected in solution. Our nucleotide-free KATNAL1 AAA monomer X-ray structure would explain how this spiral assembly might be capped and rapidly disassemble upon ADP release. The ATP-bound hexameric ring structures described by Zehr et al. (22) were detected only with a specific Glu to Gln mutation, which inactivates ATP hydrolysis. Likewise, further support for the ADP-bound left-handed spiral will require a mutation that stabilizes this transient state.

Using our two experimentally determined X-ray structures, the previously described cryo-EM models for katanin, as well as hexameric Vps4 homology models (Fig. S7), we describe a revised model for katanin MT-based oligomerization and MT severing that includes both the self-inhibited monomeric nucleotide-free and the ADP left-handed spiral states. Our model incorporates the important oligomeric conformational transition described previously by Zehr et al. (22) (Figs. 8 and S8). We suggest that p60/p80 katanin heterodimers have a poor propensity for oligomerization because of the self-inhibited
Katanin nucleotide-free state reveals disassembly mechanism

AAA conformation in our KATNAL1 nucleotide-free structure. Katanin heterodimers likely bind along the MT lattice via the MIT/Con80 interface. These katanin p60/p80 heterodimers would diffuse along MT lattices, bind ATP, and begin to assemble into right-handed open rings, recently resolved by cryo-EM (22) (Fig. 8, A and B). The assembly of these oligomers likely coincides with multiple AAA subunits binding the C-terminal β-tubulin tails, which are exposed on the MT surface (30). The formation of these spirals is likely slow and requires the effect of MT and ATP binding to each subunit to form assemblies. In the studies by Zehr et al. (22), these were stabilized by blocking ATP hydrolysis with the Glu → Gln mutant. However, previous FRET experiments suggested that this monomer to oligomer transition requires both ATP binding and MT binding (19). In vivo assembly of right-handed open rings might be promoted or activated by regulators such as ASPM (2) or patronin/CAMSAP (29), which bind the MIT/con80 complex and recruit them to MTs. Initially, the open right-handed spiral of ATP-bound protomers would thread the β-tubulin C-terminal domain into its pore through the 40-Å gap in the ring. As suggested previously by Zehr et al. (22), upon hydrolysis of the ATP bound to three adjacent protomers and release of nucleotide from one protomer, the right-handed spiral would close into a flat ring encircling the β-tubulin tail (Fig.
This transition is similar to that proposed for VPS4 (25, 28, 31, 32) and is supported by a flat ring structure of MEI-1 as determined by cryo-EM (22). However, upon hydrolysis of the ATPs associated with the remaining three protomers, we suggest that the flat hexameric ring would transition to the left-handed spiral conformation (Fig. 8D) with five ADP protomers and one nucleotide-free protomer. In contrast to the conclusions drawn by Zehr et al. (22), we suggest that katanin transitions from a closed hexamer to a left-handed spiral after ATP hydrolysis and phosphate release (Fig. S8). In this left-handed spiral state, the NBD pore loops are repositioned further away from the microtubule surface, leading to a pulling effect on the αβ-tubulin C terminus away from the MT surface. This flat ring to spiral conformational change likely induces the removal of an αβ-tubulin dimer from the MT lattice. The dissociation of ADP would result in katanin oligomers rapidly disassembling to the monomeric nucleotide-free state (Fig. 8A). Our model suggests that hexameric assembly is transient followed by one or a few tubulin removal cycles and then by rapid disassembly of katanin. Our model suggests that the hexamers rapidly disassemble and are likely not processive enzymes.

**Table 3**

<table>
<thead>
<tr>
<th>Protein</th>
<th>Source</th>
<th>Nucleotide</th>
<th>PDB ID</th>
<th>RMSD (Å)</th>
</tr>
</thead>
<tbody>
<tr>
<td>MEI-1 AAA</td>
<td>C. elegans</td>
<td>ADP</td>
<td>Present study</td>
<td>1.50</td>
</tr>
<tr>
<td>MEI-1 AAA (E293Q)</td>
<td>C. elegans</td>
<td>Apo</td>
<td>5WCB, chain A</td>
<td>2.92</td>
</tr>
<tr>
<td>MEI-1 AAA (E293Q)</td>
<td>C. elegans</td>
<td>ATP</td>
<td>5WC0, chain B</td>
<td>2.43</td>
</tr>
<tr>
<td>VPS4</td>
<td>Saccharomyces cerevisiae</td>
<td>ADP</td>
<td>5UIE, chain E</td>
<td>1.83</td>
</tr>
<tr>
<td>VPS4</td>
<td>S. cerevisiae</td>
<td>ADP-BeF₄</td>
<td>5UIE, chain A</td>
<td>2.36</td>
</tr>
<tr>
<td>VPS4</td>
<td>S. cerevisiae</td>
<td>Apo</td>
<td>5UIE, chain F</td>
<td>1.38</td>
</tr>
<tr>
<td>VPS4</td>
<td>Metallophaera sedula</td>
<td>ADP</td>
<td>4D81</td>
<td>1.41</td>
</tr>
<tr>
<td>VPS4</td>
<td>M. sedula</td>
<td>Apo</td>
<td>4D80, chain A</td>
<td>1.31</td>
</tr>
<tr>
<td>Spastin</td>
<td>Drosophila melanogaster</td>
<td>Apo</td>
<td>3BP9</td>
<td>1.41</td>
</tr>
<tr>
<td>Spastin</td>
<td>Homo sapiens</td>
<td>Apo</td>
<td>3VFD</td>
<td>1.61</td>
</tr>
<tr>
<td>Fidgetin</td>
<td>C. elegans</td>
<td>Apo</td>
<td>4L15</td>
<td>1.86</td>
</tr>
<tr>
<td>Fidgetin</td>
<td>C. elegans</td>
<td>ADP</td>
<td>4L16</td>
<td>1.81</td>
</tr>
</tbody>
</table>

The structure is superimposed onto the NBD domain (residues 207–378, Ca positions) of KATNAL1 AAA ATPase with a RMSD. | Vacular protein sorting–associated protein 4.
Zehr et al. (22) suggest that the right-handed open ring to closed hexameric ring structure transition constitutes the power stroke for pulling a β-tubulin tail from the MT lattice. This transition, however, generates a ~20-Å displacement between pore loops and collectively through sequential cycles of ATP hydrolysis, resulting in the β-tubulin C terminus being translocated 120 Å perpendicular to the MT surface. In contrast, the ADP-bound katanin AAA structure proposed in our model indicates that the transition from a right-handed spiral to a left-handed spiral will likely mobilize the pore loops more than three times this distance (~60-Å) in a single event, without processive ATP hydrolysis steps around the AAA hexameric ring. We suggest that the larger displacement followed by rapid dissociation to a self-inhibited monomeric state more likely explains how the tubulin dimer is pulled from the lattice by this dynamic AAA enzyme.

**Experimental procedures**

**Protein expression and purification of katanin complexes**

The coding regions for katanin p60 from human KATNAL1 (accession number Q9BW62.1), and *C. elegans* MEI-1 (accession number Q9BW62.1), and MEI-1 inactive mutant (E308Q) were inserted into bacterial expression vectors (pCDF-Duet-TEV) with no tag using isothermal assembly. The human KATNAL1 inactive mutant (E308Q) was generated and inserted into a bacterial expression vector (V2-H6-GST-TEV) with a His tag, a GST tag, and a TEV cleavage site. The coding regions for con-80 from *C. elegans* MEI-2 (accession number AAF62184.1) and human KATNB1 (accession number CAG33043.1) were inserted into bacterial expression vectors (pMAL-CRI-TEV vector) with an MBP (maltose-binding protein) tag and a TEV cleavage site using isothermal assembly. These constructs were confirmed by DNA sequencing.

Recombinant WT KATNAL1/B1-con80 and MEI-1/MEI-2 complexes were purified as follows. Constructs were co-transformed and expressed in BL21 bacterial strains using the T7 expression system, grown at 37 °C, and induced with 0.5 mM isopropyl thio-β-galactoside at 18 °C overnight. Cells were centrifuged and resuspended in 40 mM HEPES, pH 7.3, 300 mM NaCl, 2 mM MgCl₂, and 3 mM β-mercaptoethanol (KATN buffer). Cells were lysed using a Microfluidizer (Avastin). Extracts were clarified via centrifugation at 18,000 × g.
proteins were purified and cleaved with TEV protease while attached to the amylose resin (New England Biolabs). Proteins were further purified by ion exchange using Hitrap-SP chromatography followed by size-exclusion chromatography using a Superose 6 (10/300) column (GE Healthcare). The proteins were then used in subsequent studies as described below without freezing.

Recombinant KATNAL1 (E308Q)/B1-con80 protein was expressed in bacteria using the approach described above. The protein was purified and cleaved with TEV protease while it was attached to the amylose resin. Cleaved GST and impurities were removed by nickel-nitrilotriacetic acid, GST resin, and ion exchange using Hitrap-SP chromatography. The protein was further purified using a Superose 6 (10/300) gel filtration column. The final protein was then used in subsequent studies without freezing.

Selenomethionine-substituted (Se-MEI-1/MEI-2) katanin complex was expressed in BL21 Escherichia coli strain using a metabolic labeling strategy, where growth and expression were performed using minimal media containing all amino acids but with selenomethionine replacing Met (33). The complex was purified using the strategy described above. 1 mM DTT was used throughout the purification steps.

Biochemical analyses of oligomeric katanin assemblies

To assess the oligomeric assemblies of the KATNAL1/B1-con80 and MEI-1/MEI-2 complexes, size-exclusion chromatography was performed at 4 °C. A Superose 6 size-exclusion column or Superdex 200 (SEC) was calibrated with molecular mass standards (Bio-Rad). The protein samples were analyzed at concentrations ranging from 5 to 50 μM by mixing the complex with or without 1 mM ATP into 0.5-ml volumes and injected them into a SEC column equilibrated in KATN buffer supplemented with 1 mM ATP using an AKTA purifier system (GE Healthcare). Elution fractions (0.5 ml) were collected and analyzed via SDS-PAGE. The molar ratios for KATNAL1 or KATNAL1 (E308Q) to KATNB1-con80 and MEI-1 to MEI-2 were quantitated using densitometry. Molecular masses of various concentrations of KATNAL1/B1-con80 and/or MEI-1/MEI-2 complexes supplemented with or without 1 mM ATP were measured using SEC-MALS with KATN buffer at 25 °C. The complexes were analyzed on Superdex 200 SEC column (GE Healthcare) along with UV spectrophotometer (Agilent 1100-Series HPLC), light scattering (Wyatt Technology: miniDAWN TREOS), and the refractive index (Wyatt Technology: Optilab T-rEX). Concentration-weighed molecular masses for each peak were calculated using ASTRA v. 6 software (Wyatt Technology). The SEC-MALS fractions were collected manually for one of the concentrations (25 μM), and the compositions were checked using SDS-PAGE, confirming that complexes were intact (Fig. S2C). The remaining concentrations were performed using a Superdex 200 column at 4 °C in the presence or absence of 1 mM ATP with WT KATNAL1/B1-Con80 or E308Q mutant of KATNAL1/B1-con80, and the compositions were confirmed by SDS-PAGE (Fig. S2, A and B).

Crystallization of katanin in the ADP-bound and monomeric states

Katanin complexes (KATNAL1 (E308Q)/B1-con80 and MEI-1/MEI-2) were screened for crystallization with a com-
Katanin nucleotide-free state reveals disassembly mechanism

For cryoprotection, ADP-bound and nucleotide-free crystals were soaked for 30–60 s in corresponding reservoir solutions supplemented with 30% (v/v) glycerol or 30% (v/v) PEG 400. Crystals were mounted in a cryo-loop (Hampton Research) and then immediately flash-cooled in liquid nitrogen. Diffraction data, collected at the Argonne National Laboratory at the Advanced Photon Source microfocus 24-ID-C or 24-ID-E beamline, were processed with XDS (34) or iMosfilm (35) and Scala (36). The ADP-bound cylinder-shaped crystals were obtained from MEI-1/MEI-2 complex, and one of the best crystals was diffracted to 3.1-Å resolution in a hexagonal (P6_5) space group with cell dimensions a = 98.83 Å, b = 98.83 Å, and c = 75.08 Å. The structure was determined using phases obtained from a SAD dataset from crystals grown from selenomethionine-derived protein. Automated XDS-SHELX-PHENIX for the SAD dataset was performed by the RAPD data processing pipeline at the Synchrotron NE-CAT beamline (https://rapd.nec.aps.anl.gov). Selenium sites identified using the SHELX suite (37) were refined to a 0.65 figure of merit and then supplied to an AutoBuild module using the program RESOLVE in the PHENIX program suite (38). Automatic chain tracing

X-ray data collection and structure determination

For cryoprotection, ADP-bound and nucleotide-free crystals were soaked for 30–60 s in corresponding reservoir solutions supplemented with 30% (v/v) glycerol or 30% (v/v) PEG 400. Crystals were mounted in a cryo-loop (Hampton Research) and then immediately flash-cooled in liquid nitrogen. Diffraction data, collected at the Argonne National Laboratory at the Advanced Photon Source microfocus 24-ID-C or 24-ID-E beamline, were processed with XDS (34) or iMosfilm (35) and Scala (36).

The ADP-bound cylinder-shaped crystals were obtained from MEI-1/MEI-2 complex, and one of the best crystals was diffracted to 3.1-Å resolution in a hexagonal (P6_5) space group with cell dimensions a = 98.83 Å, b = 98.83 Å, and c = 75.08 Å. The structure was determined using phases obtained from a SAD dataset from crystals grown from selenomethionine-derived protein. Automated XDS-SHELX-PHENIX for the SAD dataset was performed by the RAPD data processing pipeline at the Synchrotron NE-CAT beamline (https://rapd.nec.aps.anl.gov). Selenium sites identified using the SHELX suite (37) were refined to a 0.65 figure of merit and then supplied to an AutoBuild module using the program RESOLVE in the PHENIX program suite (38). Automatic chain tracing
yielded several helical fragments. Manual tracing in the program COOT was used to fill the gaps (39). The density map indicated a MEI-1 AAA domain in the asymmetric unit. The N-terminal MIT domain and MEI-2 could not be identified in the density map, revealing that it underwent proteolysis during crystallization. A Matthews coefficient calculation of the solvent content supports the idea that only the MEI-1 AAA domain is contained in the crystal (40). The final model was refined using PHENIX program and torsion-angle molecular dynamics with a slow-cooling simulated annealing and TLS (Translation/Libration/Screw) schemes to $R_{\text{free}}$/R$_{\text{work}}$ (0.26/0.21). The MEI-1 AAA structure consists of residues 164–265, 270–298, 308–325, and 331–468 and an ADP molecule. Despite the presence of the nonhydrolyzable analogue AMP-PNP in the crystallization condition, only ADP was identified at the active site, suggesting that AMP-PNP underwent hydrolysis during crystallization. Data processing and refinement statistics are given in Table 2.

The best nucleotide-free rectangular crystals formed from KATNAL1 (E308Q)/B1-con80 complex crystals and diffractions to 2.4 Å. Diffraction data were collected for the best nucleotide-free rectangular crystals obtained from KATNAL1 (E308Q)/B1-con80 complex and diffracted during crystallization. Data processing and refinement statistics are given in Table 2.

The stereochemical properties of the final models, KATNAL1 (E308Q)/B1-con80 complex crystals and diffracted to 2.4 Å. Diffraction data were collected for the best nucleotide-free rectangular crystals obtained from KATNAL1 (E308Q)/B1-con80 complex crystals and diffracted during crystallization. A Matthews coefficient calculation of the solvent content supports the idea that only the MEI-1 AAA domain is contained in the crystal (40). The final model was refined using PHENIX program and torsion-angle molecular dynamics with a slow-cooling simulated annealing and TLS (Translation/Libration/Screw) schemes to $R_{\text{free}}$/R$_{\text{work}}$ (0.26/0.21). The MEI-1 AAA structure consists of residues 164–265, 270–298, 308–325, and 331–468 and an ADP molecule. Despite the presence of the nonhydrolyzable analogue AMP-PNP in the crystallization condition, only ADP was identified at the active site, suggesting that AMP-PNP underwent hydrolysis during crystallization. Data processing and refinement statistics are given in Table 2.

The best nucleotide-free rectangular crystals formed from KATNAL1 (E308Q)/B1-con80 complex crystals and diffractions to 2.4 Å. Diffraction data were collected for the best nucleotide-free rectangular crystals obtained from KATNAL1 (E308Q)/B1-con80 complex in the $P2_12_12_1$ space group to 2.4 Å resolution with unit cell dimensions $a = 40.10$, $b = 61.19$, and $c = 117.62$ Å. Phase information was determined by molecular replacement using the NBD domain (polyalanine model) of MEI-1 AAA as the search model. An unambiguous rotation and translation solution was obtained using the program PHASER (41). Gold substructures were obtained by running PHASER in its MR-SAD mode with phases from NBD polyalanine model. This phase was subjected to density modification with solvent flattening and histogram matching as implemented in the CCP4 suite (42). Automatic chain tracing using RESOLVE yielded several helical fragments of the HBD domain. Manual tracing in the program COOT was used to fill the gaps (39). The asymmetric unit contains only a KATNAL1 (E308Q) AAA domain; the N-terminal MIT domain and KATNB1 could not be identified, suggesting that these domains underwent proteolysis during crystallization and were likely lost. A Matthews coefficient calculation also supports the hypothesis that only the KATNAL1 (E308Q) AAA domain is contained in the crystal (40). The final model was refined using the PHENIX program (43) and torsion-angle molecular dynamics with a slow-cooling simulated annealing and TLS schemes to $R_{\text{free}}$/R$_{\text{work}}$ (0.25/0.22). The KATNAL1 (E308Q) AAA structure consists of residues 184–280, 285–315, 318–339, and 348–490, tetraethylene glycol, and 53 water molecules. No nucleotide was identified in the density map, despite the presence of the nonhydrolyzable analogue AMP-PNP in the crystallization condition.

The stereochemical properties of the final models, KATNAL1 (E308Q) AAA and MEI-1 AAA, were assessed using the program MolProbity (44). All structure-rendering figures were generated using UCSF Chimera (45) and PyMOL (48).

**Molecular modeling of N-terminal MIT domain and hexameric katanin assemblies**

Two different monomeric states (ATP and ADP states) of the AAA domain from the human KATNAL1 sequence (residues 184–490) were built by homology to the cryo-EM structure of the MEI-1 AAA hexameric right-handed spiral (PDB code 5WC0, ATP state, chain B (22)) and current ADP-bound MEI-1 AAA (PDB code 5B5D) structure, respectively (Fig. S7, A and B). To build a prehydrolysis ATP-state model (Fig. S7C), Ca atoms of the monomeric AAA-ATP–state models were optimally superposed to two adjacent protomers in the cryo-EM structure of the MEI-1 AAA hexameric right-handed spiral (22). This process was repeated up to a desired number of times to construct hexameric right-handed spiral assembly. The same approach was applied to get the katanin active-state model by different nucleotide states of AAA protomers (monomeric models and autoinhibited nucleotide-free structures) onto the six AAA modules of the VPS4 hexameric closed-ring structure (Fig. S7D) (25). The post-hydrolysis state model of human katanin was generated by different nucleotides states of AAA protomers (ADP state and autoinhibited nucleotide-free structures) onto the six AAA modules of the current MEI-1 AAA pseudo-hexameric left-handed spiral assembly (Fig. S7E). These models were energy minimized using the program CNS (46).

To generate a structural model of full MEI-1 P60 for mapping C. elegans mei1 meiosis mutations, a model of the N-terminal MIT domain of C. elegans was built by homology to the crystal structure of the MIT domain of the M. musculus p60N/p80C katanin complex (PDB code 5NBT (2)). The MIT domain model was generated using UCSF Chimera (45) and Modeler (47) followed by energy minimization with the CNS program (46).


**Acknowledgments**—We thank Advanced Photon Source and Drs. K. Rajashankar, J. Schuermann, and D. Neuau of the Northeastern Collaborative Access Team (NE-CAT) for the use of the 24-IDC and IDE beam lines to collect all the X-ray diffraction data for our crystallographic studies. We thank our graduate student, Brian D. Cook, for help with SEC-MALS. This work is based upon research conducted at the NE-CAT beamlines, which are funded by the National Institute of General Medical Sciences from the National Institutes of Health (P41 GM103403). The Pilatus 6M detector on 24-ID-C beam line is funded by a NIH-ORIP HEI grant (S10 RR029205). The Eiger 16M detector on 24-ID-E beam line is funded by a NIH-ORIP HEI grant (S10OD021527). This research used resources of the Advanced Photon Source, a U.S. Department of Energy (DOE) Office of Science User Facility operated for the DOE Office of Science by Argonne National Laboratory under Contract No. DE-AC02-06CH11357.

**References**

Katanin nucleotide-free state reveals disassembly mechanism


Downloaded from http://www.jbc.org at University of California, Davis on November 20, 2018
Katanin nucleotide-free state reveals disassembly mechanism


